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(51) International Patent Classification ⁵ : C12N 15/29, 15/62, 15/82 C12P 21/02, C12N 5/10 A01H 5/00	A1	(11) International Publication Number: WO 93/21320 (43) International Publication Date: 28 October 1993 (28.10.93)
(21) International Application Number: PCT/CA92/00161 (22) International Filing Date: 15 April 1992 (15.04.92) (60) Parent Application or Grant (63) Related by Continuation US 659,835 (CIP) Filed on 22 February 1991 (22.02.91) (71) Applicant (for all designated States except US): UNIVERSITY TECHNOLOGIES INTERNATIONAL, INC. [CA/CA]; ES620, 2500 University Drive, N.W., Calgary, Alberta T2N 1N4 (CA). (72) Inventor; and (75) Inventor/Applicant (for US only): MOLONEY, Maurice, M. [GB/CA]; 131 Edgehill Place, Calgary, Alberta T3A 2S4 (CA).		(74) Agent: CLARK, Geoffrey, C.; Fetherstonhaugh & Co., 456 - 409 Granville Street, Vancouver, British Columbia V6C 1H5 (CA). (81) Designated States: AT, AU, BB, BG, BR, CA, CH, CS, DE, DK, ES, FI, GB, HU, JP, KP, KR, LK, LU, MG, MN, MW, NL, NO, PL, RO, RU, SD, SE, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LU, MC, NL, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, SN, TD, TG). Published <i>With international search report.</i>
(54) Title: OIL-BODY PROTEINS AS CARRIERS OF HIGH-VALUE PEPTIDES IN PLANTS (57) Abstract Methods and compositions for use therein are described for expressing a polypeptide of interest in a seed cell as a fusion protein with an oil body protein. By this means, the fusion protein is targeted to the oil bodies of a seed cell. The oil body is easily separated from other cellular material following lysis of the seed cell, for example by using the partitioning/surface properties of the oil body. The fusion protein may be isolated for example by affinity chromatography using antibodies directed to the oil body protein. Where desired, the polypeptide of interest can be recovered by treatment of the fusion protein with for example a protease capable of recognizing a proteolytic recognition site in the oil body protein proximal to the N-terminus of the polypeptide of interest.		

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OIL-BODY PROTEINS AS CARRIERS OF
HIGH-VALUE PEPTIDES IN PLANTS

INTRODUCTION

Technical Field

10 This invention relates to a method for the
production by recombinant means of a protein of interest
which is easily purified from host cell components. The
method is exemplified by expression of the protein of
interest in plants, particularly seeds, as a chimeric
15 peptide comprising an oil-body protein and the protein of
interest.

Background

20 A variety of proteins have been expressed in
plants. However, while the general feasibility of
obtaining expression of foreign proteins in plants has been
demonstrated, obtaining purified proteins from this source
has some limitations. These limitations include the
purification step necessary to obtain pure protein
25 essentially free of plant derived materials and the
degradation that may occur in extracts prepared during the
purification procedure when the recombinant proteins
obtained are in contact with aqueous buffers.

30 Plants bearing oilseeds such as soybean,
rapeseed, sunflower and a number of other plant species
such as corn, carrot, etc., store triglycerides in their
seeds. In the plant, these triglycerides act as a source of
energy for a germinating seed and the subsequent seedling.
The triglycerides are widely used as vegetable oils in foods
35 and in food preparation and also for some industrial
applications.

 Triglycerides are immiscible with water and
partition by floating on the surface of aqueous solutions or
by forming small globules or liposomes as a suspension in

the aqueous phase. Such globules will naturally coalesce if they are not stabilized by a modified surface layer. This coalescence can result in a suspension of globules of random sizes. In seeds, when triglyceride is stored, the oil globules are actually encapsulated lipid or oil bodies normally of uniform size. Associated with the surface of these oil bodies is a half unit membrane studded with several proteins, generally referred to as oil-body proteins.

At least one class of oil-body proteins has some characteristics which are highly conserved between species. This class of oil-body proteins is referred to as an "oleosin." The hydrophilic N- and C-termini of these proteins appear to be quite divergent, whereas the lipophilic internal region (central core) appears to be highly conserved between species. The oleosins are strongly associated with the oil bodies; this strong association to the oil-bodies may, in major part, be due to the lipophilic nature of these central core. It is therefore of interest to determine whether oil body proteins such as oleosins may be useful in a method for the production of recombinant proteins by providing a means for separation of the recombinant proteins from plant derived materials.

25

Relevant Literature

The production of foreign (recombinant) peptides in plants has been investigated using a variety of approaches including transcriptional fusions using a strong constitutive plant promoter (e.g., from cauliflower mosaic virus--Sijmons et al. (1990) *Bio/Technology*, 8:217-221) and the coding of a foreign protein; transcriptional fusions with organ specific sequences (Radke et al. (1988) *Theoret. Appl. Genet.*, 75:685-694); and translational fusions which require subsequent cleavage of a recombinant protein (Vander Kerkove et al. (1989) *Bio/Technology*, 7:929-932). Foreign proteins which have been expressed in plant cells include active proteins from bacteria (Fraley et al. (1983) *Proc.*

Nat'l. Acad. Sci. USA, 80:4803-4807), animals (Misra and Gedamu (1989) *Theor. Appl. Genet.*, 78:161-168), fungi and other plant species (Fraley et al. (1983) *Proc. Nat'l. Acad. Sci. USA*, 80:4803-4807).

5 Some proteins, normally markers of integration, have been expressed in a tissue-specific manner, including some in seeds (Sen Gupta-Gopalan et al. (1985) *Proc. Nat'l. Acad. Sci. USA*, 82:3320-3324); Radke et al. (1988) *Theor. Appl. Genet.*, 75:685-694). These reports have concentrated
10 specifically on the use of seed-storage protein promoters as a means of deriving seed-specific expression. Using such a system, Vanderkerkove et al. (1989) *Bio/Technol.*, 7:929-932, expressed a high value peptide (leu-enkephalin) in seeds of *Arabidopsis thaliana* and *Brassica napus*. The yield of this
15 peptide was quite low, but demonstrates the feasibility of expression of an animal peptide hormone in plant tissues. Maize oleosin has been expressed in seed oil bodies in *Brassica napus* transformed with a maize oleosin gene. The gene was expressed under the control of regulatory elements
20 from a *Brassica* gene encoding napin, a major seed storage protein. The temporal regulation and tissue specificity of expression was reported to be correct for a napin gene promotor/terminator. See, Lee et al., *Proc. Nat'l. Acad. Sci. (USA)* (1991) 88:6181-6185.

25 The oil globules which are produced in seeds all appear to be of a similar size, indicating that they are stabilized (Huang A.H.C. (1985) in *Modern Meths. Plant Analysis*, Vol. 1:145-151 Springer-Verlag, Berlin). On closer inspection, it has been found that these are not
30 simple oil-globules, but rather oil-bodies surrounded by a membrane. These oil-bodies have been variously named by electron microscopists, oleosomes, lipid bodies and spherosomes (Gurr MI. (1980) in *The Biochemistry of Plants*, 4:205-248, Acad. Press, Orlando, Fla). The oil-bodies of a
35 few species have been studied and the general conclusion is that they are encapsulated by an unusual "half-unit" membrane comprising not a classical lipid bilayer, but rather a single amphophilic layer with hydrophobic groups on

the inside and hydrophilic groups on the outside (Huang A.H.C. (1985) in *Modern Meths. Plant Analysis*, Vol. 1:145-151 Springer-Verlag, Berlin).

Analysis of the contents of lipid bodies has demonstrated that apart from triglyceride and membranous material, there are also several polypeptides/proteins associated with the surface or lumen of the oil body (Bowman-Vance and Huang (1987) *J. Biol. Chem.*, 262:11275-11279, Murphy et al. (1989) *Biochem. J.*, 258:285-293, Taylor et al. (1990) *Planta*, 181:18-26). Oil-body proteins have been identified in a wide range of taxonomically diverse species (Moreau et al. (1980) *Plant Physiol.*, 65:1176-1180; Qu et al. (1986) *Biochem. J.*, 235:57-65) and been shown to be uniquely localized in oil-bodies and not found in organelles of vegetative tissues. In *Brassica napus* (rapeseed) there are at least three polypeptides associated with the oil-bodies of developing seeds (Taylor et al. (1990), *Planta*, 181:18-26). The numbers and sizes of oil-body associated proteins may vary from species to species. In corn, for example, there are two immunologically distinct polypeptide classes found in oil-bodies (Bowman-Vance and Huang (1988) *J. Biol. Chem.*, 263:1476-1481). Oleosins have been shown to comprise regions of alternate hydrophilicity, hydrophobicity and hydrophilicity (Bowman-Vance and Huang (1987) *J. Biol. Chem.*, 262:11275-11279). The amino acid sequences of oleosins from corn, rapeseed, and carrot have been obtained. See Qu and Huang (1990) *J. Biol. Chem.*, 265:2238-2243, Hatzopoulos et al. (1990) *Plant Cell*, 2:457-467, respectively. In an oilseed such as rapeseed, oleosin may comprise between 8% (Taylor et al. (1990) *Planta*, 181:18-26) and 20% (Murphy et al. (1989) *Biochem. J.*, 258:285-293) of total seed protein. Such a level is comparable to that found for many seed storage proteins.

Genes encoding oil-body proteins have been reported for two species, maize (*Zea mays*, Bowman-Vance and Huang (1987) *J. Biol. Chem.*, 262:11275-11279; and Qu and

Huang (1990) *J. Biol. Chem.*, 265:2238-2243) and carrot (Hatzopoulos et al. (1990) *Plant Cell*, 2:457-467).

SUMMARY OF THE INVENTION

5

Methods and compositions are provided for the production of peptides which may be easily purified from host proteins. The method includes the steps of preparing a chimeric DNA construct which includes a sequence encoding an oil-body specific sequence comprising the coding sequence of a seed-specific oil-body protein gene, or a sequence encoding at least a portion of the hydrophobic core of an oil-body protein, and a coding sequence for a peptide of interest from which an expression cassette containing the chimeric DNA construct can be prepared; transforming a host cell with the expression cassette under genomic integration conditions; and growing the resulting transgenic plant to produce seed in which the polypeptide of interest is expressed as a fusion protein with the oleosin.

The polypeptide of interest may be purified by isolating oil-bodies from the cells of the seed, and disrupting the oil-bodies so that the fusion protein is released. The oil-body protein is then easily separated from other proteins and plant derived material by phase separation. Optionally a cleavage site may be located at least one of prior to the N-terminus and after the C-terminus of the polypeptide of interest allowing the fusion polypeptide to be cleaved and separated by phase separation into its component peptides. The production system thus provides for targeting of the chimeric peptide by its oil-body protein functionality to the oil bodies which, in turn, permits rapid purification of the polypeptide of interest. This production system finds utility in the production of many peptides such as those with pharmaceutical, enzymic, rheological and adhesive properties.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1A. shows the nucleotide sequence and deduced amino-acid sequence (17 kDa protein) of an oil-body protein gene (oleosin) from *Arabidopsis thaliana*. Underlined are the direct repeats (R1 and R2) and inverted repeat (T), a CACA, TATA, TAAT and polyadenylation signals. The intron sequence is printed in lower case and a putative ABA-binding site is indicated in bold.

Fig. 1B. shows a comparison of the sequences of oil-body 16 Kd protein from carrot, an 18 Kd and 16 Kd oil-body protein from maize and a 17 Kd oil-body protein from *Arabidopsis thaliana* indicating conserved and divergent regions of the proteins; the amino acid sequences are aligned to show the conservation of sequence in the central region of the proteins.

Fig. 2. shows constructs used for the fusion of oil-body protein genes with genes encoding foreign peptides. IA is a C-terminal fusion of a desired peptide to OBP; IB is an N-terminal fusion of a desired peptide to OBP; II is an internal fusion of a desired peptide within OBP; and III is an inter-dimer translational fusion of desired peptide enclosed between two substantially complete oil body protein targeting sequences. In the upper portion of Figure (A) are shown the DNA constructs used for translational fusions of desired peptides to oil-body proteins. In the lower portion of Figure (B) are shown the configurations of the gene products, shown on the upper portion of the translation and the delivery to the oil bodies. The key to the figure is as follows: bottom left-top right hatched box represents an OBP promoter or other seed specific promoter; bottom right-top left hatched box represents a desired peptide coding sequence; open box represents an oil-body protein coding sequence or synthetic targeting sequence based on OBP conserved motifs; vertical-horizontal hatched box represents a gene terminator containing a polyadenylation signal; hatched circle

represents a protease recognition motif; corkscrew line represents a native C- or N-terminal of OBP.

Fig. 3. shows a detailed arrangement for construction of a C-terminal fusion. Shown is the arrangement is a collagenase recognition motif coding sequence as a linker in the fusion of a typical oil-body protein gene and a fusion peptide, to be linked here using an *NcoI*, for cloning and expression in plants.

Fig. 4. shows schematically the process of construction of fusion peptide vectors, their introduction into plants and subsequent extraction and assay of the desired recombinant peptide.

Figure 5 shows a schematic representation of the construction of pCGOBPILT. The broken line box represents an oleosin promoter; the top left-bottom right hatched box represents an oleosin coding sequence; the horizontal-vertical hatched box represents an intron; the dotted box represents a 3' non-translated sequence; and the widely-spaced top left-bottom right hatched box represents an interleukin-1- β sequence equipped with a sequence encoding a protease cleavage site (Factor Xa or thrombin immediately upstream).

Figure 6 shows the design of oligonucleotide GVR11. In Figure 3A represents the 3' coding sequence of the *A. thaliana* oleosin, translationally fused to the factor Xa/IL-1- β coding sequence followed by a TAA stop codon. For future cloning purposes, a *PvuI* and *SalI* restriction enzyme recognition site are included. The creation of a *PvuI* restriction site resulted in the additional coding sequence for an alanine (ala). Underlined are the restriction enzyme recognition sequences. Overlined are the *A. thaliana* oleosin sequences and the factor Xa recognition sequence. The actual cleavage site is indicated with an asterisk (*). In Figure 3B, the sequence of GVR11 is shown. In order to make a fusion with the *A. thaliana* oleosin, the primer GVR11 needs to be a sequence complementary to the top strand.

Figure 7 shows the nucleotide sequence of OBPILT. Underlined is the sequence encoding IL-1- β ; the sequence encoding the factor Xa recognition site is indicated in bold. The nopaline synthase terminator sequence is indicated in lower case letters.

DESCRIPTION OF THE SPECIFIC EMBODIMENTS

10 In accordance with the subject invention, methods and compositions are provided for production of peptides which are easily purified. The subject method includes the steps of preparing an expression cassette containing DNA sequences encoding a sufficient portion of an oil body
15 specific sequence, such as an oleosin, to provide for targeting to an oil body and the peptide of interest; transforming the expression cassette into a plant cell host; generating a transgenic plant and growing it to produce seed in which the chimeric protein is expressed and translocated
20 to the oil bodies. The chimeric peptide comprises the peptide of interest and an oil body protein such as an oleosin. The peptide of interest generally is a foreign peptide normally not expressed in seeds or found on the oil-body. The use of an oil-body protein as a carrier or
25 targeting means provides a simple mechanism to obtain purification of the foreign protein. The chimeric protein is separated away from the bulk of cellular protein in a single step (such as centrifugation or flotation); the protein is also protected from degradation during extraction
30 as the separation also removes non-specific proteases from contact with the oil-bodies. The gene encoding the foreign peptide may be derived from any source, including plant, bacterial, fungal or animal source. Desirably, the chimeric peptide will contain sequences which allow for cleavage of
35 the peptide of interest from the oleosin. The method may be employed to express a variety of peptides which are then easily purified.

Targeting a foreign, recombinant protein to the oil-body imparts several advantages, including the following. The protein can be separated from the bulk of cellular contents after cell lysis by centrifugation. The oil-body fraction will float on the surface of the extract. The protein can optionally be provided with a peptide linker containing a protease recognition site. This permits release of the peptide from the oil-body. The protein can be introduced into a recombinant polypeptide in such a way that it is within a lipophilic conserved region. This results in the internalization of the recombinant peptide into the oil-body, thus protecting it from protease attack.

The expression cassette generally will include in the 5'-3' direction of transcription, a transcriptional and translational regulatory region capable of expression in developing seed, typified by the promoter and upstream regions associated with an oil body protein, which will provide for expression of the chimeric protein in seed, a DNA sequence encoding a chimeric peptide comprising an amino acid sequence to provide an oil body targeting means and a protein of interest, and a transcriptional and translational termination region functional in plants. One or more introns may also be present.

The oil-body specific sequence finds analogy in fragments of oil-body proteins, particularly oleosins. The oil-body specific sequence may be the same as that of a sequence obtainable from an oil-body protein, or which has sufficient homology to provide for the desired targeting of a protein of interest to an oil body. By "obtainable" is intended an amino acid sequence which may be natural, synthetic or a combination, sufficiently similar to a native oil body protein amino acid sequence to provide the desired targeting. Of particular interest is the central hydrophobic domain of oil body proteins which appears to be highly conserved among different plant species, and fragments thereof and homologous sequences at the amino acid level.

10

The deduced amino acid sequence for an *Arabidopsis thaliana* oil-body protein is as follows:

```

                    10                20
M-N-G-R-D-R-D-Q-Y-Q-M-S-G-R-G-S-D-Y-S-K-
5                    30                40
S-R-Q-I-A-K-A-A-T-A-V-T-A-G-G-S-L-L-V-L-
                    50                60
L-S-L-T-L-V-G-T-V-I-A-L-T-V-A-T-P-L-L-V-
                    70                80
10 I-S-S-T-I-L-V-P-A-L-I-T-V-A-L-L-I-T-G-S-
                    90                100
L-S-S-G-G-F-G-I-A-A-I-T-V-F-S-W-I-Y-K*Y-
                    110               120
L-L-I-E-H-P-Q-G-S-D-K-L-D-S-A-R-M-K-L-G-
15                    130               140
S-K-A-Q-D-L-K-D-R-A-Q-Y-Y-G-Q-Q-H-T-G-W-
                    150
E-H-D-R-D-R-T-R-G-G-Q-H-T-T

```

20 Amino acids from about 25-101 comprise the central hydrophobic domain.

Of particular interest as a targeting means for some applications are oil-body specific sequences or fragments thereof of the following formula which provide
25 for targeting to an oil body:

```

pp1- aa25- aa26- V - V- T - L- aa21- P-
                    A   A   A   T
30 aa34- G- G- aa36- L- L- aa39- L- aa41-
                    M
35 G - I - aa44- L- aa46- aa47- T - L - I -
S   L                               S   V   V
40 aa51- L- aa53- V- A- T- P- L - aa59- L -
                    V   V
L - F- S- P- V - L- V- P- A- A - L - aa73-
I   -   I   L   I

```

SUBSTITUTE SHEET

aa⁷⁴ - aa⁷⁵ - aa⁷⁶ - aa⁷⁷ - aa⁷⁸ - G- F- L-

5 G L
S - S- aa⁸⁷ - G- V - aa⁸⁹ - aa⁹⁰ - L - S -
T I I T

10 aa⁹³ aa⁹⁴ - S - aa⁹⁶ - aa⁹⁷ - aa⁹⁸ - aa⁹⁹ -
 T
 aa¹⁰⁰ - aa¹⁰¹ - pp²

15 wherein:

pp¹ and pp² are the same or different, and may be the same as or different from a natural oil-body protein, usually different; they may be hydrogens, indicating the terminal portion of the indicated polypeptide or may be polypeptides having a total of up to 1000 amino acids, more usually of up to about 500 amino acids, and may have a total of as few as 1 amino acid, or may individually or separately be polypeptides of from 1-100 amino acids, more usually from about 1-75 amino acids, more particularly from about 5-50 amino acids; these polypeptides will have specific applications in modifying a specifically described sequence for a predetermined purpose;

aa²⁵ may be any amino acid, particularly a neutral aliphatic amino acid, generally of 3-6 carbon atoms, more particularly leucine or alanine;

aa²⁶ is a neutral aliphatic amino acid, particularly alanine or an hydroxy substituted amino acid of from 3-4 carbon atoms, particularly threonine or a basic amino acid of from 5-6 carbon atoms, particularly lysine;

35 aa³¹ is a neutral unsubstituted aliphatic amino acid of from 3-6 carbon atoms, particularly alanine, valine or leucine or an aromatic unsubstituted amino acid, particularly phenylalanine;

40 aa³³ is a neutral unsubstituted aliphatic amino acid of from 3-6 carbon atoms, particularly alanine, valine or leucine or an oxy-substituted aliphatic amino acid, particularly threonine;

- aa³⁶ is a neutral aliphatic unsubstituted amino acid of from 3-5 carbon atoms, particularly leucine or a neutral aliphatic oxy-substituted amino acid of from 3-4 carbon atoms, particularly threonine or serine;
- 5 aa³⁷ is a neutral unsubstituted amino acid, particularly leucine or a thio-substituted amino acid, particularly methionine;
- aa³⁹ is a neutral aliphatic unsubstituted amino acid, particularly valine or an aromatic unsubstituted amino acid, particularly phenylalanine;
- 10 aa¹ is a neutral aliphatic unsubstituted or oxy-substituted amino acid, particularly alanine, leucine or serine;
- aa⁴⁴ is a neutral aliphatic unsubstituted or oxy-substituted amino acid, particularly alanine, isoleucine or threonine;
- 15 aa⁴⁶ is a neutral aliphatic unsubstituted amino acid or an oxy-substituted amino acid, particularly alanine, valine or threonine;
- 20 aa⁴⁷ is a neutral aliphatic unsubstituted amino acid, particularly glycine or alanine;
- aa⁵⁹ is a neutral aliphatic or aromatic unsubstituted amino acid, particularly leucine or phenylalanine;
- 25 aa⁷⁶ is a neutral aliphatic unsubstituted or thio-substituted amino acid, particularly alanine, leucine or methionine;
- aa⁷⁸ is a neutral aliphatic unsubstituted amino acid, particularly alanine or a neutral aliphatic amino acid having a thio- or an oxy-substitution, particularly methionine or threonine;
- 30 aa⁸³ is a neutral aliphatic unsubstituted or oxy-substituted amino acid, particularly glycine, serine or threonine;
- 35 aa⁹² is a neutral aliphatic amino acid with a oxy-substitution, particularly serine or threonine;

aa⁹⁶ is a neutral aliphatic thio-substituted amino acid or a neutral aromatic heterocyclic amino acid, particularly tryptophan;

aa⁹⁷ is a neutral aliphatic unsubstituted or
5 thio-substituted amino acid, particularly valine, leucine, isoleucine or methionine;

aa⁹⁸ is a neutral aliphatic unsubstituted amino acid or an aromatic oxy-substituted amino acid, particularly alanine, leucine or tyrosine;

10 aa⁹⁹ may be any amino acid;

aa¹⁰⁰ is an oxy-substituted amino acid, either aliphatic or aromatic, particularly tyrosine or threonine;

aa¹⁰¹ is a neutral unsubstituted aliphatic or aromatic amino acid, particularly alanine, leucine or
15 phenylalanine.

Of particular interest as a source of DNA encoding sequences capable of providing for targeting to an oil body protein are oil-body protein genes obtainable from *Arabidopsis* or *Brassica napus* which provide for expression
20 of the protein of interest in seed (See Taylor et al. (1990) *Planta*, 181:18-26). The necessary regions and amino-acid sequences to provide targeting ability to the oil body appear to be the highly hydrophobic central region of oil body proteins.

25 To identify other oil body protein genes having the desired characteristics, where an oil body protein has been or is isolated, the protein may be partially sequenced, so that a probe may be designed for identifying mRNA. Such a probe is particularly valuable if it is designed to target
30 the coding region of the central hydrophobic domain which is highly conserved among diverse species of plants. In consequence, a DNA or RNA probe for this region may be particularly useful for identifying coding sequences of oil body proteins from other plant species. To further enhance
35 the concentration of the mRNA, cDNA may be prepared and the cDNA subtracted with mRNA or cDNA from non-oil body producing cells. The residual cDNA may then be used for

probing the genome for complementary sequences, using an appropriate library prepared from plant cells. Sequences which hybridize the cDNA under stringent conditions may then be isolated.

5 In some instances, as described above, using an oil body protein gene probe (conserved region), a probe may be employed directly for screening a cDNA genomic library and identifying sequences which hybridize to the probe. The isolation may also be performed by a standard immunological
10 screening technique of a seed-specific cDNA expression library. Antibodies may be obtained readily for oil-body proteins using the purification procedure and antibody preparation protocol described by Taylor et al. (*Planta*, (1990) 181, 18-26). cDNA expression library screening
15 using antibodies is performed essentially using the techniques of Huynh et al. (1985, in *DNA Cloning*, Vol. 1, a *Practical Approach*, ed. D.M. Glover, IRL Press, pp. 49-78). Confirmation of sequence is facilitated by the high conservation found in the central hydrophobic region (see
20 Fig. 1). DNA sequencing by the method of Sanger et al. (*Proc. Natl. Acad. Sci. USA*, (1977) 74:5463-5467) or Maxam and Gilbert (1980, *Meth. Enzymol.*, (1980) 65:497-560) may be performed on all putative clones and homology searches performed. Homology of sequences encoding the central
25 hydrophobic domain is normally $\geq 70\%$, both at the amino-acid and nucleotide level between diverse species. If an antibody is available, confirmation of sequence identity may also be performed by hybrid-select and translation experiments from seed mRNA preparations as described by
30 Sambrook et al. (*Molecular Cloning*, (1990) 2nd Ed., Cold Spring Harbor Press, pp. 8-49 to 8-51).

 cDNA clones made from seed can be screened using cDNA probes made from the conserved coding regions of any available oil body protein gene (e.g., Bowman-Vance and
35 Huang (*J. Biol. Chem.*, (1987) 262:11275-11279). Clones are selected which have more intense hybridization with seed DNAs as compared to seedling cDNAs. The screening is repeated to identify a particular cDNA associated with oil

bodies of developing seeds using direct antibody screening or hybrid-select and translation. The mRNA complementary to the specific cDNA is absent in other tissues which are tested. The cDNA is then used for screening a genomic
5 library and a fragment selected which hybridizes to the subject cDNA.

To obtain expression of the chimeric gene in seed a transcriptional initiation regulatory region and translational initiation regulatory region of untranslated
10 5' sequences, "ribosome binding sites", responsible for binding mRNA to ribosomes and translational initiation obtainable from any gene preferentially expressed in seed may be used. Examples of such genes include seed storage proteins such as from napin (Josefsson et al., *J. Biol.*
15 *Chem.*, (1987) 262:12196-12201; Scofield S.R. and Crouch M.L. *J. Biol. Chem.* (1987) 262:12202-12208). Preferably, the region is obtainable from an oil body protein (oil-body proteins from *Arabidopsis*, carrot (Hatzopoulos et al., *supra*) or maize (Huang et al. 1987 and 1990 *supra*). The
20 region generally comprises at least 100 bp 5' to the translational start of the structural gene coding sequence, up to 2.5 kb 5' to the same translational start. It is preferred that all of the transcriptional and translational functional elements of the initiation control region are
25 derived from or obtainable from the same gene. By "obtainable" is intended a DNA sequence sufficiently similar to that of a native sequence to provide for the desired specificity of transcription of the DNA sequence encoding the chimeric protein. It includes natural and synthetic
30 sequences and may be a combination of synthetic and natural sequences.

The transcription level should be sufficient to provide an amount of RNA capable of resulting in a modified seed. By "modified seed" is meant seed having a detectably
35 different phenotype from a seed of a non-transformed plant of the same species, for example one not having the expression cassette in question in its genome. Various changes in phenotype are of interest. These changes include

over-expression of oil-body protein or OBP-accumulation on the oil body or in the cytoplasm of the resultant chimeric protein.

The polypeptide of interest may be any protein and includes for example, an enzyme, an anticoagulant, a neuropeptide, a hormone, or adhesive precursor. Examples of proteins include interleukin-1- β , the anticoagulant Hirudin, the enzyme β -glucuronidase or a single-chain antibody comprising a translational fusion of the V_H or V_L chains of an immunoglobulin. The DNA sequence encoding the polypeptide of interest may be synthetic, naturally derived, or combinations thereof. Depending upon the nature or source of the DNA encoding the polypeptide of interest, it may be desirable to synthesize the DNA sequence with plant preferred codons. The plant preferred codons may be determined from the codons of highest frequency in the proteins expressed in the largest amount in the particular plant species of interest as a host plant.

The termination region which is employed will be primarily one of convenience, since in many cases termination regions appear to be relatively interchangeable. The termination region may be native with the transcriptional initiation region, may be native with the DNA sequence encoding the polypeptide of interest, or may be derived from another source. Convenient termination regions are available from the Ti-plasmid of *A. tumefaciens*, such as the octopine synthase and nopaline synthase termination regions.

Ligation of the DNA sequence encoding the targeting sequence to the gene encoding the peptide of interest may take place in various ways including terminal fusions, internal fusions, and polymeric fusions. In all cases, the fusions are made so as not to interrupt the reading frame of the oil-body protein and so as to avoid any translational stop signals in or near the junctions. The different types of terminal and internal fusions are shown in Fig. 2 along with a representation of their configurations *in vivo*.

In all the cases described, the ligation of the gene encoding the peptide preferably would include a linker encoding a protease target motif. This would permit the release of the peptide once extracted as a fusion protein. Potential cleavage sites which could be employed are recognition motifs for thrombin (leu-val-pro-arg-gly) (Fujikawa et al., Biochemistry (1972) 11:4892-4899), of factor Xa (phe-glu-gly-arg-aa.) (Nagai et al., Proc. Nat'l Acad. Sci. USA, (1985) 82:7252-7255) or collagenase (pro-leu-gly-pro) (Scholtissek and Grosse Gene (1988) 62:55-64).

By appropriate manipulations, such as restriction, chewing back or filling in overhangs to provide blunt ends, ligation of linkers, or the like, complementary ends of the fragments can be provided for joining and ligation. In carrying out the various steps, cloning is employed, so as to amplify the amount of DNA and to allow for analyzing the DNA to ensure that the operations have occurred in proper manner. A wide variety of cloning vectors are available in *E. coli* and a marker which allows for selection of the transformed cells. Illustrative vectors include pBR332, pUC series, M13mp series, pACYC184, etc. Thus, the sequence may be inserted into the vector at an appropriate restriction site(s), the resulting plasmid used to transform the *E. coli* host, the *E. coli* grown in an appropriate nutrient medium and the cells harvested and lysed and the plasmid recovered. Analysis may involve sequence analysis, restriction analysis, electrophoresis, or the like. After each manipulation the DNA sequence to be used in the final construct may be restricted and joined to the next sequence, where each of the partial constructs may be cloned in the same or different plasmids.

A variety of techniques are available for the introduction of DNA into plant cell host. For example, the chimeric DNA constructs may be introduced into host cells obtained from dicotyledenous plants, such as tobacco, and oleaginous species, such as *Brassica napus* using standard *Agrobacterium* vectors by a transformation protocol such as

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- that described by Moloney et al. *Plant Cell Rep.*, (1989) 8:238-242 or Hinchee et al. *Bio/Technol.*, (1988) 6:915-922; or other techniques known to those skilled in the art. For example, the use of T-DNA for transformation of plant cells
- 5 has received extensive study and is amply described in EPA Serial No. 120,516; Hoekema, In: *The Binary Plant Vector System* Offset-drukkerij Kanters B.V., Alblasterdam, 1985, Chapter V, Knauf, et al., *Genetic Analysis of Host Range Expression by Agrobacterium*, In: *Molecular Genetics of the*
- 10 *Bacteria*. Plant interaction, Puhler, A. ed., Springer-Verlag, NY, 1983, p. 245, and An et al., *EMBO J.* (1985), 4:277-284. Conveniently, explants may be cultivated with *A. tumefaciens* or *A. rhizogenes* to allow for transfer of the transcription construct to the plant cells.
- 15 Following transformation using *Agrobacteria* the plant cells are dispersed in an appropriate selective medium for selection, grown to callus, shoots grown and plantlets regenerated from the callus by growing in rooting medium. The *Agrobacterium* host will contain a plasmid having the *vir*
- 20 genes necessary for transfer of the T-DNA to the plant cells and may or may not have T-DNA. For injection and electroporation, (see below) disarmed Ti-plasmids (lacking the tumor genes, particularly the T-DNA region) may be introduced into the plant cell.
- 25 The use of non-*Agrobacterium* techniques permits the use of the constructs described herein to obtain transformation and expression in a wide variety of monocotyledonous and dicotyledonous plants. These techniques are especially useful for species that are
- 30 intractable in an *Agrobacterium* transformation system. Other techniques for gene transfer include biolistics (Sanford, *Trends in Biotech.* (1988) 6:299-302), electroporation (Fromm et al. (1985) *Proc. Nat'l. Acad. Sci. USA*, 82:5824-5828; Riggs and Bates (1986), *Proc. Nat'l. Acad. Sci. (USA)* 83 5602-5606 or PEG-mediated DNA
- 35 uptake (Potrykus et al. (1985) *Mol. Gen. Genet.*, 199:169-177).

As a host cell, cells from any of a number of seed bearing plants may be employed in which the cells are derived from plant parts such as stem, leaf, root, or seed or reproductive structures according to the species. The cells may be isolated cells or plant parts, for example, leaf discs. In a specific application, such as to *Brassica napus*, the host cells generally will be derived from cotyledonary petioles as described by Moloney et al. *Plant Cell Rep.*, (1989) 8:238-242). Other examples using commercial oil seeds include cotyledon transformation in soybean explants (Hinchey et al. *Biotechnology*, (1988) 6:915-922) and stem transformation of cotton (Umbeck et al. *Biotechnology*, (1981) 5:263-266).

Following transformation, the cells, for example as leaf discs, are grown in selective medium. Once shoots begin to emerge, they are excised and placed onto rooting medium. After sufficient roots have formed, the plants are transferred to soil. Putative transformed plants are then tested for presence of a marker. Southern blotting is performed on genomic DNA using an appropriate probe, for example an *A. thaliana* oleosin gene, to show that integration of the desired sequences into the host cell genome has occurred.

The expression cassette will normally be joined to a marker for selection in plant cells. Conveniently, the marker may be resistance to a herbicide, particularly an antibiotic, such as kanamycin, G418, bleomycin, hygromycin, chloramphenicol, or the like. The particular marker employed will be one which will allow for selection of transformed cells as compared to cells lacking the DNA which has been introduced.

The fusion peptide in the expression cassette constructed as described above, expresses at least preferentially in developing seeds. Accordingly, transformed plants grown in accordance with conventional ways, are allowed to set seed. See, for example, McCormick et al. *Plant Cell Reports* (1986) 5:81-84. Northern blotting can be carried out using an appropriate gene probe with RNA

isolated from tissue in which transcription is expected to occur such as a seed embryo. The size of the transcripts can then be compared with the predicted size for the fusion protein transcript.

5 Oil-body proteins are then isolated from the seed and analyses performed to determine that the fusion peptide has been expressed. Analyses can be for example by PAGE. The fusion peptide can be detected using an antibody to the oleosin portion of the fusion peptide. The size of the
10 fusion peptide obtained can then be compared with predicted size of the fusion protein.

Two or more generations of transgene plants may be grown and either pollinated with the same transformed strain or different strains, identifying the resulting hybrid
15 having the desired phenotypic characteristic, to ensure that the subject phenotypic characteristic is stably maintained and inherited and then seeds harvested for isolation of the peptide of interest or for use to provide seeds with the new phenotypic property.

20 The desired protein can be extracted from seed that is homo- or heterozygous for the introduced trait by a variety of techniques, including use of an aqueous, buffered extraction medium and a means of grinding, breaking, pulverizing or otherwise disrupting the cells of
25 the seeds. The extracted seeds can then be separated (for example, by centrifugation or sedimentation of the brei) into three fractions: a sediment or insoluble pellet, an aqueous supernatant, and a buoyant "scum" comprising seed storage lipid and oil bodies. These oil bodies contain both
30 native oil-body proteins and chimeric oil body proteins, the latter containing the foreign peptide. The oil-bodies are separated from the water-soluble proteins and re-suspended in aqueous buffer.

If a linker comprising a protease recognition
35 motif has been included in the expression cassette, to the resuspension buffer is added a protease specific for the recognition motif produced by translation of the linker sequence. This releases the required peptide into the

aqueous phase. A second centrifugation step will now re-float the processed oil-bodies with their attached proteins and leave an aqueous solution of the required peptide. The desired peptide may be precipitated,
5 chemically modified or lyophilized according to its properties and desired applications

In certain applications it may not be necessary to remove the chimeric protein from the oil-body protein. Such an application would include cases where the fusion peptide
10 includes an enzyme which is tolerant to N or C-terminal fusions and retains its activity; such enzymes could be used without further cleavage and purification. The chimeric enzyme-OBP would be contacted with substrate as a fusion protein. It is also possible, if desired, to purify
15 the enzyme - OBP fusion protein using an immunoaffinity column comprising an immobilized high titre antibody against the OBP (see, for example, Taylor et al., (1990) supra).

Other uses for the subject invention are as follows. OBP's comprise a high percentage of total seed
20 protein, thus it is possible to enrich the seed for certain desirable properties such as high-lysine, high methionine, and the like, simply by making the fusion protein rich in the amino-acid(s) of interest could find utility of particular interest is the modification of grains and
25 cereals which are used as either directly or indirectly as food sources for livestock, including cattle, poultry, and humans. It may be possible to include, as the fusion peptide, an enzyme which may assist in subsequent processing of the oil or meal in conventional oilseed crushing and
30 extraction, for example inclusion of a thermostable lipid-modifying enzyme which would remain active at the elevated crushing temperatures used to process seed and thus add value to the extracted triglyceride or protein product. Other uses of the fusion protein to include use to improve
35 the agronomic health of the crop. For example, an insecticidal protein or a portion of an immunoglobulin specific for an agronomic pest such as a fungal cell wall or

membrane, could be coupled to the oil body protein thus reducing attack of the seed by a particular plant pest.

The following examples are offered by way of illustration and not by limitation.

5

EXPERIMENTAL

Example 1

Expression of terminal fusions of foreign peptides with oil-body proteins

10

A. C-terminal fusions

A genomic clone of an oil-body protein gene containing at least 100 bp 5' to the translational start is
15 cloned into a plasmid vehicle capable of replication in a suitable bacterial host (e.g., pUC or pBR322 in *E. coli*). A restriction site is located in the region encoding the hydrophilic C-terminal portion of gene. In a 19 kDa OBP, this region stretches typically from codons 125 to the end
20 of the clone. The ideal restriction site is unique, but this is not absolutely essential. If no convenient restriction site is located in this region, one may be introduced by the site-directed mutagenesis procedure of Kunkel *Proc. Nat'l. Acad. Sci. USA*, (1985) 82:488-492. The
25 only major restriction on the introduction of this site is that it must be placed 5' to the translational stop signal of the OBP clone.

With this mutated clone in place, a synthetic oligonucleotide adapter may be produced which contains
30 coding sequence for a protease recognition site such as Pro-Leu-Gly-Pro or a multimer thereof. This is the recognition site for the protease collagenase. The adaptor would be synthesized in such a way as to provide: a 4-base overhang at the 5' end compatible with the restriction site
35 at the 3' end of the OBP clone, a 4-base overhang at the 3' end of the adaptor to facilitate ligation to the foreign peptide coding sequence and additional bases, if needed, to ensure no frame shifts in the transition between the OBP

coding sequence, the protease recognition site and the foreign peptide coding sequence. A typical arrangement for such a fusion is shown in Fig. 3. The example shown here uses an existing *Xho*I site near the stop codon of a carrot OBP (Hatzopoulos et al. *Plant Cell*, (1990) 2:457-467). This is digested and may be ligated with an adapter constructed from the two oligonucleotides described. This adapter will form a perfect *Xho*I overhang at an end and will not disrupt the translational frame. The other end forms an *Nco*I overhang which is arbitrarily chosen (any six-base cutter will suffice), but which encloses an ATG from the desired foreign peptide.

The final ligation product will contain an almost complete OBP gene, coding sequence for collagenase recognition motif and the desired peptide coding region all in a single reading frame. This tripartite fragment is cloned into an *Agrobacterium* binary plasmid (Bevan *Nucl. Acid Res.*, (1984) 12:8711-8721) such as is widely used to transfer foreign DNA into plants (Fraley et al. *Proc. Nat'l. Acad. Sci. USA*, (1983) 80:4803-4807) and this is used to transform oilseed plants such as rapeseed using the method of Moloney et al. *Plant Cell Rep.*, (1989) 8:238-242) or similar procedure. Transgenic plants may be recovered from this transformation experiment and these are grown to flowering. The plants then set seed by self-fertilization.

The seeds are allowed to reach maturity (60-80 days) and then are harvested and ground in aqueous extraction buffer (Taylor et al. *Planta*, (1990) 181:18-26). The slurry is centrifuged at 5000 xg for 20 min. and will give a surface scum. This scum is again recovered and suspended by vigorous shaking in a collagenase assay buffer (Scholtissek and Grosse, *Gene* (1988) 62:55-64). Five units of collagenase are added and the suspension is incubated with shaking for 4 h. After this time, the suspension is once again centrifuged at 5000 xg for 20 min. The surface scum is removed and the protein content of the aqueous phase is analyzed by SDS-Poly Acrylamide Gel Electrophoresis. If

a band of approximately the size of the required peptide is found, the protein may be precipitated using ammonium sulfate, concentrated using ultrafiltration or lyophilized.

5 B. N-terminal fusions

The hydrophilic N-terminal end of oil-body proteins permits the fusion of peptides to the N-terminal while still assuring that the foreign peptide would be retained on the outer surface of the oil body. The configuration of such fusions is shown in Fig. 2IB.

This configuration can be constructed from similar starting materials as used for C-terminal fusions, but requires the identification of a convenient restriction site close to the translational start of the oil-body protein gene. A convenient site may be created in many oil-body protein genes without any alteration in coding sequence by the introduction of a single base change just 5' to the first 'ATG'. In oil body proteins thus far studied, the second amino acid is alanine whose codon begins with a "G". The context of the sequences is shown below:

A-C transition here yields NcoI site

↓
 3' . . . TC TCA ACA ATG GCA . . . Carrot OBP
 3' . . . CG GCA GCA ATG GCG . . . Maize 18KDa OBP

A single base change at the adenine prior to the 'ATG' would yield in both cases . . . CCATGG . . . which is an NcoI site. Thus, modification of this base using the site-directed mutagenesis protocol of Kunkel (*Proc. Nat'l. Acad. Sci. USA*, (1985) 82:488-492) will prepare this clone for use assuming no other NcoI sites in the sequence.

The coding sequence for the foreign peptide may require preparation which will allow its ligation directly into the NcoI site. This may typically require a single or two-base modification by site-directed mutagenesis (Kunkel, 1985, *supra*) to generate an NcoI site around the

translational start of the foreign peptide. This peptide is then excised from its cloning vehicle using *Nco*I and a second enzyme which cuts close to the translational stop of the target. Again, using the methods described above, a second convenient site can be introduced by site-directed mutagenesis. It has been suggested by Qu and Huang (1990, *supra*) that the N-terminal methionine might be removed during processing of the protein *in vivo* and that the alanine immediately downstream of this might be acylated. To account for this possibility, it may be necessary to retain the Met-Ala sequence at the N-terminal end of the protein. This is easily accomplished using a variety of strategies which introduce a convenient restriction site into the coding sequence in or after the Ala codon. For example, by site-directed mutagenesis, the sequences could be modified as follows:

3' ... TC TCA ACA ATG GCA GAA CGA GGC ACT TAT

mutate to

3' ... TC TCA ACA ATG GCA TGC CGA GGC GCC TAT

SphI

NarI

This change of a single codon would introduce a *Sph*I site into the coding sequence. A second change, which could be introduced during the same round of mutagenesis would convert two bases in codon 6 to yield GGC GCC, an *Nar*I site. This mutated gene could then be opened with *Sph*I and *Nar*I to give a directional cloning cut which would eliminate three codons. Into this site could be introduced an adaptor containing a 3' overhang with the sequence CATG ... (compatible with *Sph*I) and a GC 5' overhang at the opposite end. The precise sequence of this adapter is shown below:

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Sph1

Narl

n times

5

CTACG

CCG	CTC	GGT	CCG	GG
GGC	GAG	CCA	GGC	CCGC

This adapter would recreate both the *Sph*I and *Nar*I restriction sites which would be used for diagnostic purposes. The *Sph*I site could now be used to open the plasmid and clone in-frame a DNA fragment enclosing the sequence for a useful peptide. Orientation of cloning could then be analyzed by cutting at any asymmetrically placed site and *Nar*I of the plasmid.

The resultant constructs from these N-terminal fusions would be typical of the examples IB of Figure 2. They would contain an OBP promoter sequence, an in-frame fusion in the first few codons of the OBP gene of a high value peptide coding sequence with its own ATG as start signal if necessary and the remainder of the OBP gene and terminator.

This modified gene is introduced into a binary *Agrobacterium* plasmid (Bevan, (1984), supra) and mobilized into *Agrobacterium*. Transformations are performed as described above. Recovery of the high value peptide from seeds is performed as described for 'C-terminal fusions.'

C. Internal translational fusions

A third type of fusion involves the placing of a high value peptide coding sequence internally to the coding sequence of the OBP. This type of fusion requires the same strategy as in N-terminal fusions, but may only be functional with modifications in regions of low conservation, as it is believed that regions of high conservation in these OBPs are essential for targeting of the mature protein.

The key difference in this kind of fusion is the necessity for *flanking* collagenase recognition sites for the release of the protein. This means that in place of the standard collagenase linker/adaptor systems thus far

described, it is necessary to have a linker with the following form:

5
 Cohesive end 1 n times n times Restriction site end 2
 CCG CTC GGT CCG GGC GAG CCA GGC CCG CTC GGT GGC GAG CCA GGC
 10

Cohesive ends 1 and 2 would be used to clone the adapter into an OBP clone in a directional manner. The nested restriction site is then used to introduce the high value peptide coding sequence flanked by appropriate restriction sites or linkers. Orientation is checked by the use of an asymmetrically placed restriction site in the high-value peptide coding sequence and one of the two restriction sites flanking the coding sequence of the collagenase recognition motif.

Mobilization of these constructs to *Agrobacterium* plasmids and then to plants is identical to the previously described procedure. Recovery of the high-value protein from the seeds of transgenic plants is somewhat different in that after the oil-bodies have been isolated and washed, it may be necessary to delipidate the oil-bodies in order to access the collagenase recognition sites which could be hidden inside the oil-body in the lipid phase. This step may reduce certain advantages of using oil-body proteins as carriers, but may on the other hand be very convenient for protein sequences which are labile in aqueous media or in plant cytoplasm.

D. Inter-dimer translational fusions

It is possible to create a construct in which the entire coding sequence of the OBP is repeated. A dimeric protein produced from this construct may still contain all the necessary factors for targeting the OBP to the oil-body. Such a construct would contain a promoter region, an entire or near-complete open reading frame for an OBP but excluding

the translational stop and then an entire open-reading frame of a second OBP, this time equipped with a translational 'stop' and a terminator region.

In the construction of this chimeric gene a pair
5 of dissimilar restriction sites are either found or created at the region of the junction of the two copies. These sites are used to enable the introduction of a linker such as is described above for internal translational fusions. The linker contains not only sets of collagenase recognition
10 motifs, but also an internal restriction site in which to nest a sequence encoding a high value protein. The form of this construct is shown in Fig. 2 III. Mobilization of this construct to *Agrobacterium* and then to plants is exactly as above. Recovery of the high value protein from seeds of the
15 transformed plants would be performed using the same procedure as described for C-terminal fusions above.

Example 2

Strategy for the cloning and expression of 20 Interleukin-1- β (IL-1- β) as a fusion with oleosins in plants

A. Cloning and sequencing of an *Arabidopsis thaliana* oleosin gene

25 A *Brassica napus* oleosin gene (Murphy et al, (1991) *Biochim Biophys Acta* 1088: 86-94) was used to screen a genomic library of *A. thaliana* (cv. Columbia) in EMBL3A (Stratagene). The screening resulted in the isolation of a EMBL3A clone (λ 2.1) containing a 15 kb genomic fragment
30 from *A. thaliana*. The oleosin was mapped within a 6.6 kb KpnI insert, within this 15 kb fragment (Fig. 5). A 1.8 kb NcoI/KpnI fragment containing the oleosin gene was end filled and subcloned in the SmaI site of RFM13mp19. The 1.8 kb insert was digested with convenient restriction enzymes
35 and subcloned in M13mp19 for sequencing. The 1800 bp sequence of the *A. thaliana* oleosin gene is presented in Fig. 1a. All the cloning procedures were carried out according to Sambrook et al., (1989) (Molecular Cloning: A

laboratory manual 2nd ed. Cold Spring Harbor Laboratory Press.)

B. Design of an oligonucleotide encoding IL-1- β

- 5 IL-1- β consists of 9 amino acids (aa); val-gln-gly-glu-glu-ser-asn-asp-lys (Antoni et al., (1986) *J. Immunol.* 137:3201-3204). The protease factor Xa can cleave a protein sequence which contains a aa sequence ile-glu-gly-arg. Cleavage takes place after the aa arg. Based on these
- 10 sequences an oligonucleotide was designed (GVR11, fig. 5), which contains in addition to the IL-1- β coding sequence, the coding sequence for the factor Xa cleavage site, and 18 nucleotides of the 3' coding region of the *A. thaliana* oleosin (base position 742-759). The IL-1- β coding sequence
- 15 was designed using optimal codon usage for the *B. napus* and *A. thaliana* oleosin (Table 1).

Table 1.

Codon usage of *A. thaliana* and *B. napus oleosin*²

TTT	phe	F	5	TCT	ser	S	8	TAT	tyr	Y	6	TCT	cys	C	-
TTC	phe	F	3	TCC	ser	S	10	TAC	tyr	Y	11	TGC	cys	C	-
TTA	leu	L	-	TCC	ser	S	2	TAA	och	Z	-	TGA	opa	Z	-
TTG	leu	L	4	TGA	ser	S	-	TAG	amb	Z	-	TGG	trp	W	2
CTT	leu	L	10	CCT	pro	P	3	CAT	his	H	4	CGT	arg	R	6
CTC	leu	L	11	CCC	pro	P	1	CAC	his	H	6	CGC	arg	R	-
CTA	leu	L	-	CCA	pro	P	4	CAA	gln	Q	4	CGA	arg	R	1
CTG	leu	L	6	CCG	pro	P	2	CAG	gln	Q	17	CGG	arg	R	-
ATT	ile	I	7	ACT	thr	T	11	AAT	asn	N	-	AGT	ser	S	5
ATC	ile	I	13	ACC	thr	T	14	AAC	asn	N	1	AGC	ser	S	3
ATA	ile	I	3	ACA	thr	Y	7	AAA	lys	K	5	AGA	arg	R	5
ATG	met	M	11	ACG	thr	T	5	AAG	lys	K	10	AGG	arg	R	3
GTT	val	V	11	GCT	ala	A	17	GAT	asp	D	8	GGT	gly	G	0
GTC	val	V	9	GCC	ala	A	2	GAC	asp	D	19	GGC	gly	G	9
GTA	val	V	-	GCA	ala	A	10	GAA	glu	E	2	GGA	gly	G	14
GTG	val	V	-	GCG	ala	A	2	GAG	gly	E	2	GGG	gly	G	5

1 See Figure 1A.

2 Lee and Huang (1991). Plant Physical 96: 1395-1397.

C. Creation of an *A. thaliana* oleosin-IL-1- β fusion

Based on the sequence:

5' CACACCAGGAAGTCTCTGGTAAGC 3'

5 (base position: -838 to -814), oligonucleotide GVR10

5' CACTGCAGGAAGTCTCTGGTAAGC 3'

was designed. GVR10 contains a *Pst*I restriction site (underlined) to facilitate cloning. The polymerase chain reaction (PCR) was used to amplify the region between GVR10 and GVR11. The reaction mixture contained: 16 μ l dNTPs (1.25 mM), 10 μ l 10X PCR buffer (100 mM Tris-HCL pH 8.3, 500 mM KCL, 15 mM MgCl₂, 0.1% (w/v) gelatin), 5 μ l GVR11 (20 μ M) 1 μ l Taq DNA polymerase (1 u/ μ l) and 64 μ l H₂O. The reaction was carried out for 30 cycles. Each cycle consisted of 1 minute denaturing at 92°C, 1 minute annealing at 45°C and 3 minutes extension at 72°C. The PCR reaction yielded a single fragment of 1652 nucleotides.

D. Cloning of the *A. thaliana* oleosin-IL-1- β (OBPIL) fusion

A 5' *Sal*I-nopaline synthase (nos) terminator-*Eco*RI 3' sequence was isolated from pBI121 (Clontech laboratories) and cloned into the *Sal*/*Eco*RI sites of pUC19. The plasmid was called pTerm. The 1652 bp fragment (described in C.) was isolated and digested with the restriction enzymes *Pst*I and *Sal*I. This fragment was cloned in pTerm. The resulting plasmid was called pUCOBPILT (fig. 5). This plasmid was digested with *Eco*RI and *Pst*I and resulted in the digested pUC19 vector and the *Eco*RI-A. *thaliana* oleosin-IL-1- β -nos -*Pst*I fusion *Pst*I (OBPILT). The complete sequence of OBPILT is shown in fig. 7. OBPILT was subcloned in the *Eco*RI/*Pst*I sites of pBluescript+. This plasmid (pBIOBPILT) was digested with *Pst*I and *Hind*III and the *Pst*I-OBPILT-*Hind*III fragment was subcloned in a binary *Agrobacterium* plasmid (Bin 19) (Bevan, M., (1984) *Nucl. Acid. Res.* 12: 8711-8721) containing a selection marker (neomycin phosphotransferase and *Pst*I-*Hind*III unique sites. The resulting plasmid was called pCJOBPILT. A schematic

representation of the cloning procedure is shown in Fig. 5. For descriptions of various binary plasmids, see, pGA642 or 645; An et al. (1985) EMBO. J. 4 277-288 or pCGN1558 or 1559; MacBride and Summerfeldt (1990) *Plant. Molec. Biol.* 14 269-276.

F. Transformation of pCGBPILT into *Agrobacterium* strain EHA101

A single EHA101 colony (Hood et al., (1986) *J. Bact.* 168:1291-1301) was used to inoculate 5 ml of LB+100 μ g/ml kanamycin. This culture was grown for 48 hours at 28°C. This 5 ml culture was used to inoculate 500 ml of LB+100 μ g/ml kanamycin. This culture was grown at 28°C until the culture reached a density of OD600=0.5 (approx. 4 hours). The cells were spun down (10 min, 5000 x g) and resuspended in 500 ml of sterile H₂O (repeated 2x). The cells were spun again and resuspended in 3 ml sterile H₂O, containing 10% glycerol. 40 μ l of the cells were aliquoted in Eppendorf tubes and either directly used for electroporation, or stored at -80°C for future use. Electroporation was carried out according to Bower et al., *Nucl. Acid. Res.* (1988) 16 6127-6145. The pulse generator was set to the 25 μ F capacitor, 2.5 kV and 200 ohm in parallel with the sample chamber.

G. Transformation of *Nicotiana tabacum* (tobacco) with pCGBPILT

The EHA 101 containing pCGBPILT was used to transform tobacco leaf discs. Eight to ten centimeter long tobacco leaves were taken from greenhouse grown plants, sterilized in 70% ethanol for 20 sec. and then in 10% bleach (such as Javex) for 8 min. The leaves were then rinsed 6 times with sterile water. The leaf edges as well as the midrib were excised from the leaves and the remaining lamina was sectioned into 5x7 mm squares or discs of 5 mm diameter. About 30 leaf discs were collected and placed into a small petri dish. The *Agrobacterium* solution was then poured over the tobacco discs and incubation occurred for 9 minutes.

The leaf pieces were then blotted on sterile Whatman filter paper and placed, abaxial side down, onto media I (MS, 3% sucrose and 2 mg/l 2,4-D). Co-cultivation proceeded for the following 48 hours. At this point the leaf discs were transferred to selection media (MD, 3% sucrose, 2.5 mg/l Ba, 0.1 mg/l NAA, 500 mg/l carbenicillin, and 100 mg/l kanamycin) where they remained for the next 3-4 weeks. Once shoots began to emerge they were excised and placed onto rooting media (MS, 3% sucrose, 0.1 mg/l NAA, 500 mg/l carbenicillin, and 50 mg/l kanamycin). After sufficient roots had been formed, the tobacco plants were transferred to soil.

H. Transformation of *B. napus* with pCGBPILT

The transformation of *B. napus* was carried out according to Moloney et al., (1989) *Plant Cell Rep*, 8:238-242, which disclosure is incorporated herein by reference.

Transformation Procedure

Single colonies of *Agrobacterium tumefaciens* strain EHA 101 containing the binary plasmid were grown overnight at 28°C in AB medium. A 50 µl sample of this suspension was grown overnight at 28°C in 5 ml of MG/L broth supplemented with appropriate antibiotics. This bacterial suspension was pelleted by centrifugation for 15 min. at 10,000 x g then resuspended in 10 ml of MS medium containing 3% sucrose and at pH 5.8. A thin film of this suspension was used to cover the base of a 5 cm petri dish. Individual excised cotyledons were taken from the plates described above and the cut surface of their petioles was immersed into this bacterial suspension for a few seconds. They were immediately returned to the same MS plates from which they had been taken. The cotyledons were co-cultivated with the *Agrobacterium* for 72 h. No feeder layers were employed.

After co-cultivation, the cotyledons were transferred to regeneration medium comprising MS medium supplemented with 20 µM benzyladenine, 3% sucrose, 0.7% phytagar, pH 5.8 and 500 mg/l carbenicillin (Pyopen,

Ayerst) and 15 mg/l kanamycin sulphate (Boehringer-Mannheim). Again the petioles were carefully embedded in the agar to a depth of 2 mm. Plating density was maintained at 10 explants per plate. Higher densities
5 reduce regeneration frequency.

Selection and Plant Regeneration

The explants were maintained on regeneration
10 medium under light and temperature conditions specified above for 2-3 weeks. During this time many shoots appeared on over half the explants with relatively little callus formation. Some of these shoots undergo bleaching by the fourth week of culture. The remaining green shoots were
15 subcultured onto shoot elongation medium which was the same as regeneration medium minus the benzyladenine. One or two weeks on this medium permitted the establishment of apical dominance from the shoot clusters formed. The shoots so derived were transferred to "rooting" medium containing MS
20 medium, 3% sucrose, 2mg/l indole butyric acid, 0.7% phytagar and 500 mg/l carbenicillin. No kanamycin was used at this stage as it was found that more rapid root establishment occurred without the selection agent while very few
"escapes" actually succeeded in rooting after the two
25 rounds of selection on regeneration and shoot elongation medium.

I. Stable integration of OBPIIT in the tobacco and B. napus genomes

30 Putative transformed plants were tested for neomycin phosphotransferase activity. Genomic DNA from plants showing this activity was isolated. Southern blotting was performed in order to demonstrate that the sequences between the T-DNA borders (OBPIIT and neomycin
35 phosphotransferase gene) were stably integrated in to the genomes of B. napus and tobacco. The tobacco Southern was probed with the A. thaliana oleosin gene, and the neomycin

phosphotransferase gene. The *B. napus* Southern was probed with the neomycin phosphotransferase gene.

J. Expression of the oleosin-IL-1- β fusion in tobacco plants

RNA was isolated from developing embryos obtained from transformed and untransformed plants. Northern blotting was carried out using the *A. thaliana* oleosin as a gene probe. In all the tested transformed plants a 850 nt transcript could be detected. The size of these transcripts correspond to the expected size of the oleosin-IL-1- β mRNA. These transcripts could not be detected in the untransformed plants.

K. Accumulation of the oleosin-IL-1- β protein

Oil-body proteins were isolated from transformed tobacco seeds (Holbrook et al., (1991) *Plant Physical* 97:1051-1058. PAGE was performed and the protein were transferred from the gel to PVDF membranes. An antibody, which was raised against a 22 kDa oleosin of *B. napus*, was used to detect the oleosin-IL-1- β fusion in the tobacco seeds. This antibody recognizes all the major oleosins in *B. napus* and *A. thaliana*. In addition, this antibody recognizes the tobacco oleosins. Tobacco oleosins have different sizes from the *A. thaliana* and *B. napus* oleosins. In the transformed tobacco seeds the anti-22 kDa antibody recognized a 20 kDa-protein, which was not present in the untransformed tobacco seed. The predicted size of the oleosin-IL-1- β fusion is 20.1 kDa. A summary of the results is shown in Table 2.

By expressing a peptide of interest conjugated to an oil body protein, or a sufficient portion thereof to provide for getting to the oil bodies, the peptide of interest can be easily purified so as to be substantially free of other cellular components. The fusion protein can be cleaved following purification or may be used without cleavage. The subject methods and compositions provide a fast, simple method for purifying a polypeptide of interest.

All publications and patent applications mentioned in this specification are herein incorporated by reference to the same extent as if each individual
5 publication or patent application was specifically and individually indicated to be incorporated by reference.

The invention now being fully described, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without
10 departing from the spirit or scope of the appended claims.

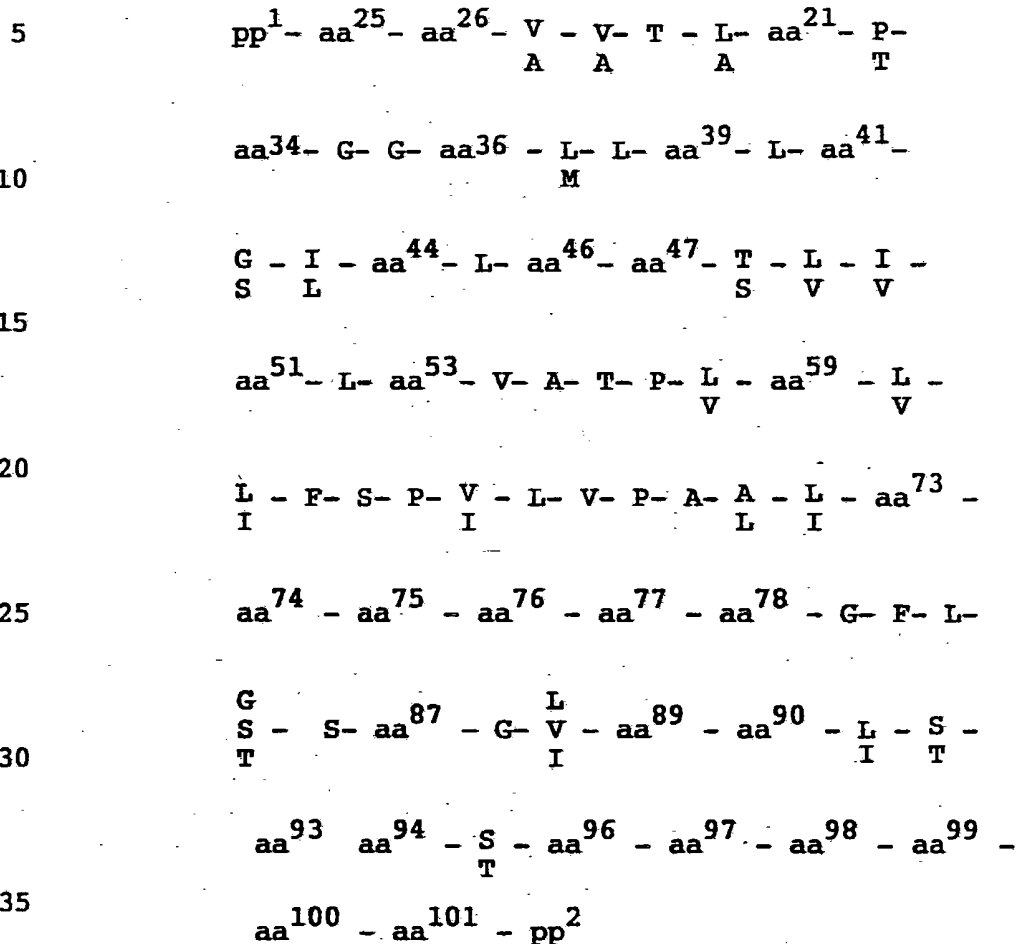
Table 2.
Summary of expression results¹

	Stable integration Southern Blotting	Expression of Oleosin -IL-1b Northern Blotting	Detection of 20kDa protein Western Blotting
B. napus:			
not transformed	-	NT	NT
transformant #12	+	NT	NT
Tobacco:			
not transformed	-	+	+
transformant A	+	+	+
transformant B	+	+	+
transformant G	+	+	+
transformant L	+	+	+

¹ Stable integration, determined as described in I, supra. Expression of the oleosin-IL-1-b fusion as described in J, supra. Detection of a putative oleosin -IL-1-b fusion protein was as described in K, supra. NT=Not Tested

WHAT IS CLAIMED IS:

1. A polypeptide, characterized as capable of targeting an oil body, and having the formula



with the proviso that said peptide is other than the full length naturally occurring 16Kd oleosin from carrot or the 18 Kd or 16 Kd oleosin from maize.

2. The polypeptide according to Claim 1, wherein at least one of PP¹ and PP² comprises a polypeptide of interest.

3. The polypeptide according to Claim 1, wherein at least one of PP¹ and PP² comprises an antigenic amino acid sequence to provide an immunogen.

4. A polypeptide characterized as capable of targeting an oil body selected from the group consisting of:

- (a) a peptide comprising at least eight consecutive amino acids included in the following amino acid sequence:

	10	20
	M-N-G-R-D-R-D-Q-Y-Q-M-S-G-R-G-S-D-Y-S-K-	
	30	40
	S-R-Q-I-A-K-A-A-T-A-V-T-A-G-G-S-L-L-V-L-	
10	50	60
	L-S-L-T-L-V-G-T-V-I-A-L-T-V-A-T-P-L-L-V-	
	70	80
	I-S-S-T-I-L-V-P-A-L-I-T-V-A-L-L-I-T-G-S-	
	90	100
15	L-S-S-G-G-F-G-I-A-A-I-T-V-F-S-W-I-Y-K*Y-	
	110	120
	L-L-I-E-H-P-Q-G-S-D-K-L-D-S-A-R-M-K-L-G-	
	130	140
	S-K-A-Q-D-L-K-D-R-A-Q-Y-Y-G-Q-Q-H-T-G-W-	
20	150	
	E-H-D-R-D-R-T-R-G-G-Q-H-T-T; and	

- (b) a peptide that is encoded by a DNA sequence identified by means of an oligonucleotide probe designed based upon said amino acid sequence in (a) or a fragment thereof, with the proviso that said peptide is other than the full length naturally occurring 16Kd oleosin from carrot or the 18 Kd or 16 Kd oleosin from maize.

5. The polypeptide according to Claim 4, wherein said peptide in (a) comprises at least twelve consecutive amino acids included in the following amino acid sequence:

40

30

40

A-K-A-A-T-A-V-T-A-G-G-S-L-L-V-L-

50

60

L-S-L-T-L-V-G-T-V-I-A-L-T-V-A-T-P-L-L-V-

5

70

80

I-S-S-T-I-L-V-P-A-L-I-T-V-A-L-L-I-T-G-S-

90

100

L-S-S-G-G-F-G-I-A-A-I-T-V-F-S-W-I-Y-K*Y-

101

10

L.

6. A DNA construct comprising a DNA sequence encoding (a) an oleosin or a portion thereof sufficient to provide for targeting to an oil body and (b) a polypeptide of interest.

7. The DNA construct according to Claim 6, further comprising:

vector DNA containing at least one regulatory sequence operatively associated with said DNA sequence which is capable of directing replication of said DNA in a host cell.

8. The DNA construct according to Claim 7, wherein said regulatory sequence is further capable of directing expression of said DNA in a host cell.

9. The DNA construct according to Claim 6, wherein said DNA is cDNA.

30

10. An expression cassette comprising: as components, in the direction of transcription, a first DNA sequence comprising a sufficient portion of the region 5' to the translational start site of a gene expressed in seed to provide for expression of a DNA sequence in seed; a second DNA sequence encoding an oleosin or a sufficient portion thereof to provide for targeting to an oil body, said second DNA sequence including at least one

natural or synthetic restriction site; and a translational and transcriptional termination region; wherein said components are operably linked and expression of said second DNA sequence is regulated by said first DNA sequence.

11. The expression cassette according to Claim 10, wherein said first DNA sequence is from a gene expressed in a cereal or grain seed cell.

12. The expression cassette according to Claim 10, wherein the genome of Aradopsis thaliana comprises at least one of said first DNA sequence and said second DNA sequence.

13. The expression cassette according to Claim 10, wherein said second DNA sequence encodes a peptide from the group consisting of:

(a) a peptide comprising at least eight amino acids of and up to the full sequence of the following amino acid sequence:

	10	20
	M-N-G-R-D-R-D-Q-Y-Q-M-S-G-R-G-S-D-Y-S-K-	
	30	40
25	S-R-Q-I-A-K-A-A-T-A-V-T-A-G-G-S-L-L-V-L-	
	50	60
	L-S-L-T-L-V-G-T-V-I-A-L-T-V-A-T-P-L-L-V-	
	70	80
	I-S-S-T-I-L-V-P-A-L-I-T-V-A-L-L-I-T-G-S-	
30	90	100
	L-S-S-G-G-F-G-I-A-A-I-T-V-F-S-W-I-Y-K*Y-	
	110	120
	L-L-I-E-H-P-Q-G-S-D-K-L-D-S-A-R-M-K-L-G-	
	130	140
35	S-K-A-Q-D-L-K-D-R-A-Q-Y-Y-G-Q-Q-H-T-G-W-	
	150	
	E-H-D-R-D-R-T-R-G-G-Q-H-T-T; and	

(b) a peptide that is encoded by a DNA sequence identified by means of an oligonucleotide probe designed based upon said amino acid sequence in (a) or a fragment thereof.

5

14. An expression cassette comprising:
an oil body protein (OBP) gene which includes a sufficient portion of the region 5' to the translational start site to provide for expression of said gene in a seed
10 cell and which includes at least one restriction site between just 5' to the codon for the initiating methionine and 5' to the translational stop signal of said OBP gene.

15. The expression cassette according to Claim
15 14, further comprising a DNA sequence encoding a polypeptide of interest inserted into said restriction site in reading frame with said OBP gene.

16. The expression cassette according to Claim
20 14, wherein said restriction site is a synthetic restriction site.

17. The expression cassette according to Claim
14, further comprising an oligonucleotide adapter coding for
25 a protease recognition site inserted into said restriction site.

18. The expression cassette according to Claim
17, wherein said protease is collagenase.

30

19. An expression cassette comprising:
a first DNA sequence encoding a polypeptide of interest inserted in reading frame into an oil body protein (OBP) gene which includes a sufficient portion of the
35 regulatory region 5' to the translational start site of said OBP gene to provide for expression of said gene in seed, wherein said sequence is inserted at a site in said gene so as to be expressed under said regulatory region.

20. The expression cassette according to Claim 19, further comprising a second DNA sequence encoding a protease recognition site 5' to said first DNA sequence, wherein said second DNA sequence is in reading frame with said first DNA sequence and said OBP gene.

21. A method for obtaining expression of a polypeptide of interest in seed, said method comprising:
transforming a host plant cell with an expression cassette under genomic integration conditions, wherein said expression cassette comprises as components, in the direction of transcription, a first DNA sequence comprising a sufficient portion of the region 5' to the translational start site of a gene expressed in seed to provide for expression of a DNA sequence in seed; a second DNA sequence encoding an oleosin or a sufficient portion thereof to provide for targeting to an oil body, said second DNA sequence including at least one natural or synthetic restriction site into which is inserted in reading frame a third DNA sequence encoding a polypeptide of interest; and a translational and transcriptional termination region; wherein said components are operably linked and expression of said second DNA sequence is regulated by said first DNA sequence to provide for expression in seed.

22. The method according to Claim 21, wherein the genome of *Thaliana aridopsis* comprises at least one of said first and said second DNA sequence.

23. A method for obtaining expression of a polypeptide of interest in seed, said method comprising:
transforming a host plant cell with a DNA construct under genomic integration conditions, wherein said DNA construct comprises a first DNA sequence encoding a polypeptide of interest inserted in reading frame into an oil body protein (OBP) gene which includes a sufficient portion of the regulatory region 5' to the translational

start site of said OBP gene to provide for expression of said gene in seed, wherein said sequence is inserted at a site in said gene so as to be expressed under said regulatory region, whereby said DNA construct becomes

5 integrated into the genome of said plant cell; and

growing said plant to produce seed whereby said polypeptide of interest is expressed as a fusion protein with the expression product of said OBP gene.

10 24. The method according to Claim 23, further comprising:

isolating said fusion protein from oil bodies in cells of said seed.

15 25. The method according to Claim 24, wherein said isolating comprises:

lysing cells of said seed to release said oil bodies; and

20 disrupting said oil bodies whereby said fusion polypeptide is released.

26. The method according to Claim 25, wherein said isolating further comprises:

25 contacting said fusion polypeptide with a protease capable of recognizing a protease recognition site in said fusion polypeptide located prior to the N-terminus of said polypeptide of interest.

30 27. The method according to Claim 26, further comprising:

prior to said contacting, binding said fusion protein to a solid support comprising an antibody capable of binding to the expression product of said OBP gene.

35 28. A method for obtaining a purified polypeptide of interest, said method comprising:

transforming a host plant cell with a DNA construct under genomic integration conditions, wherein

said DNA construct comprises a first DNA sequence encoding a polypeptide of interest inserted in reading frame into an oil body protein (OBP) gene which includes a sufficient portion of the regulatory region 5' to the translational start site of said OBP gene to provide for expression of said gene in seed, wherein said sequence is inserted at a site in said gene so that expression of said DNA sequence is controlled by said regulatory region, whereby said DNA construct becomes integrated into the genome of said plant cell;

growing said plant to produce seed whereby said polypeptide of interest is expressed as a fusion protein with the expression product of said OBP gene;
isolating oil bodies from the cells of said seed;
disrupting said oil bodies whereby said fusion protein is released; and
purifying said polypeptide of interest.

29. The method according to Claim 28, wherein said polypeptide of interest is other than a polypeptide encoded by a plant genome.

30. The method according to Claim 28, wherein said polypeptide of interest is other than a polypeptide naturally present in an oil body.

31. The method according to Claim 30, wherein said isolating comprises:
collecting an oil-body fraction following lysis of cells from said seed.

32. A plant cell comprising:
an expression cassette according to any one of Claims 10, 14, or 19.

33. A plant comprising cells containing an expression cassette according to any one of Claims 10, 14, or 19.

34. A plant expressing a polypeptide of interest in seed, obtained according to the method of Claim 23.

5 35. Seed comprising an expression cassette according to any one of Claims 10, 14, or 19.

36. Seed expressing a polypeptide of interest, obtained according to the method of Claim 23.

10

37. A method for obtaining a polypeptide of interest in an oil body, said method comprising:

expressing said polypeptide in seed as a fusion protein with an oleosin, or a sufficient portion thereof to
15 provide targeting to said oil body.

NcoI

-867 CCATGGCTATACCCACCTCGGTCTTGGTCACACAGGAACCTCTCTGGTAAGCTAGCTCCACTCCCGGAGAAACACCGGCGCCAAATTCG
 -777 COGAATTGCTGACCTGAACGGAACATCATCGTCGGTCTTGGCGGATTCGCGGAAOATGGGTGAGTCCAGCTTGGGCTTGAGGACGAGAC
 -687 CCGAATCGAGCTCTGTTGAAAGCTTGTTCATTGGGATTTGTATACGGAGATTGGTCTCGAGAGGTTGAGGAAAGGACAAATGGGTTTG
 -597 GCTCTGGAGAAAGAGAGTGGCGCTTTTAGAGAGAGAAATTGAGAGGTTTAGAGAGAGATGCGGCGCGGATGACGGGAGGAGAGACGACGAGG
 -507 ACCTGCATTATCAAAGCAGIGACGTGGTGAATTTGGAACCTTTAAAGAGGCAGATAGAITTATTATTGTATCCATTCTTCTCATTTGTTTC
 -417 TAGAATGTCCGGGAAACAAATTTTAAACCTAAATCTTAAATTTTCTAAATTTTGTGTCCTAAATAGTGGATATGTGGCCGTATAGAGGAAT
 -327 CTATTGAAGGCCAAACCCCATACTGACGAGGCCAAAGGTTCTTTCGCTTTTATGTTTCGGTTCGATGCCAACGCCACATCTCTGAGCTA
 -237 GGCAAAAACAAACCGTGTCTTTGAATAGACTCTCTCTGTTAAACACATGACGCGGCTGCATGGTGACGCCATTAAACACGTGGCCTACAAAT
 -147 GCATGATGTCTCCATTGACACGCTGACTTCTCGTCTCCTTTCTTAATATATCTAACAAACACTCCTACCTCTTCCAAATATATACACATC
 -57 TTTTGTGATCAATCTCTCATTCATAAATCTCATTTCTCTAGTAAACAAGAACAAAAATGGCGGATACAGCTAGAGGAACCCCATCAGAT
 34 I I G R D Q Y P M M G R D R D Q Y Q M S G R G S D Y S K S R
 ATCATCGGACAGACACAGTACCCGATGATGGCCGAGACCGAGACAGTACCAGATGTCCGACGAGGATCTGACTACTCCAAAGTCTAGG
 124 Q I A K A A T A V T A G G S L L V L S L T L V G T V A A L
 CAGATTGCTAAAGCTGCACTGTGTCACAGCTGGTGGTCCCTCTCTCTCCAGCCCTTACCCCTTGTGGAACTGTCTCATAGCTTTG
 214 T V A T P L L V I F S P I L V P A L I T V A L L I T G F L S
 ACTGTTGCAACACCTCTGCTCGTTATCTTCAGCCCAATCCTTGTCCCGCTCTCATCACAGTTGCACCTCCTCATCACCCTTCTCTTCC
 304 S G G F G I A A I T V F S W I Y K
 TCTGGAGGGTTTGGCATTCGGCTATAACCGTTTCTCTTGGATTACAAgtaagcacacatttatcatcttacttataaattttgtgca
 394 atatgtgcatgtgttgagccagtagctttggatcaattttttgtcgaaatacaaaatgtaacaataaagaaattgcaaatcttagg
 484 gaacatttggtaactaaatacgaatttgacctagcttagctgtaattgtctgtgtatcatctataggtataaagtcttggtatga
 574 tacctattgattgtgaatagGTACCGCAACGGGAGAGCACCCACAGGGATCAGACAAAGTTGGACAGTGCAGGATGAAGTTGGGAAGCAAA
 A Q D ; L D R A Q U U G Q Q J T G G E J D R D R T R G G Q H
 664 GCTCAGGATCTGAAAGACAGAGCTCAGTACTACGGACAGCAACATACTGGTGGGGAACATGACCGTGACCGTACTCGTGGTGGCCAGCAC
 T T *
 754 ACTACTTAAGTTACCCCACTGATGTCATCGTTCATAGTCCCAATAACTCCAATGTCGGGGAGTTAGTTTATGAGGAATAAAAGTGTTTTGAAT
 844 TTGATCAGGGGGAGATAATAAAGCCGAGTTTGAATCTTTTGTGTTAAGTAATGTTTATGTGTGTTTCTATATATGTTGTCAAAATGGTACC

FIG. 1A.

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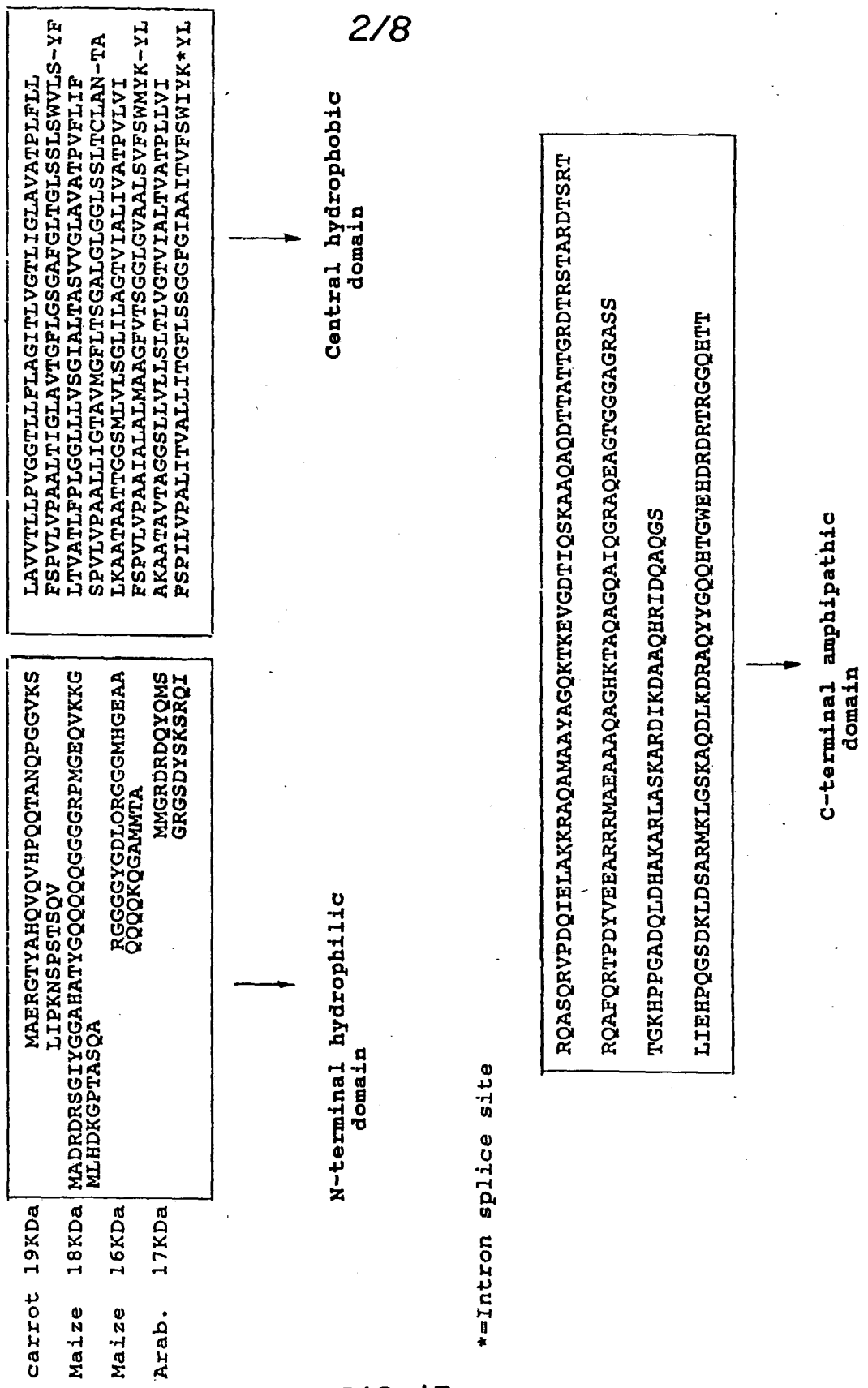


FIG. 1B.
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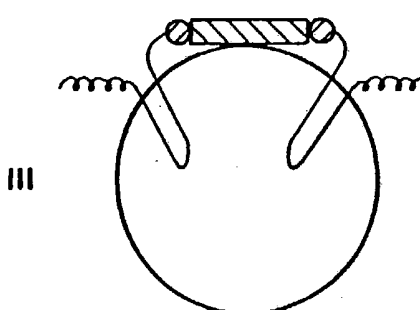
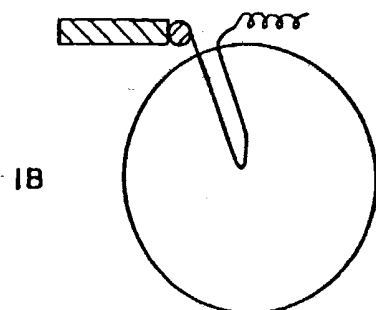
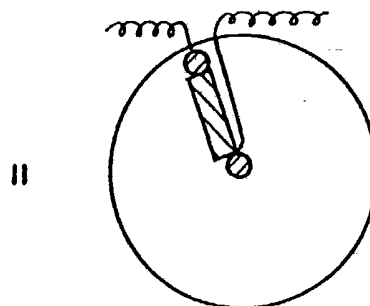
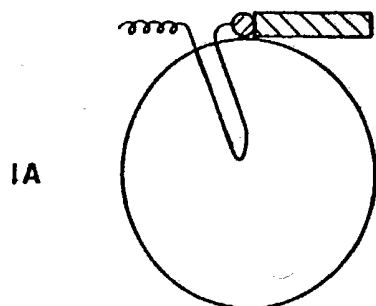
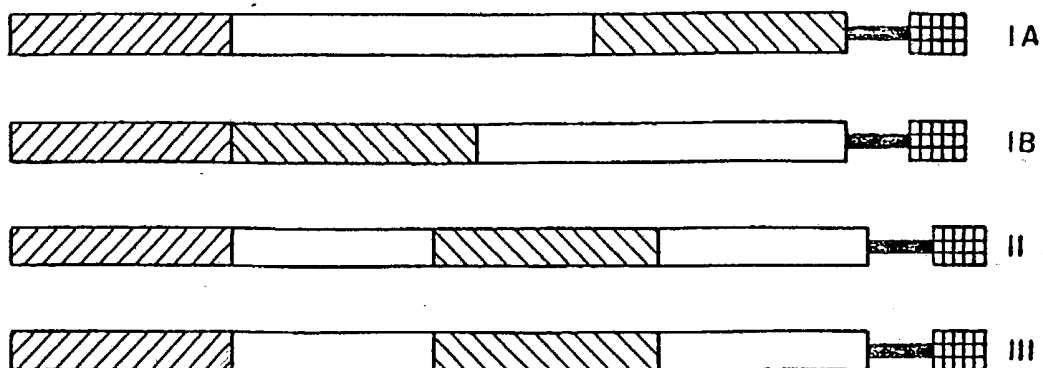


FIG. 2.

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Xho I

....AGC ACT GCT CGA GAC ACT TCA AGG ACT....Carrot OBP
 (see Hatzopoulos et
 Ser Thr Ala Arg Asp Thr Ser Arg Thr.... al. 1990)

	Xho I		n times	Nco I
3'..AGC ACT GC		TCGA	CCG CTC GGT CCG GC	
TCG TGA CGAGCT			GGC GAG CCA GGC CGGTAC	

Pro Leu Gly Pro

Collagenase recognition motif

FIG. 3.

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Construct fusion gene using protease (eg collagenase) target
motif coding linkers clone in E. coli compatible plasmid



Insert construct into wide host range replicon containing T-DNA
borders (i.e. Agrobacterium binary vector)



Transform plant-cells using leaf, stem, cotyledonary
or petiole explants



Regenerate transgenic plants



Allow to set seed



Grind seed in aqueous extraction buffer
(Taylor et al, 1990)



Centrifuge or otherwise separate oil-bodies



Suspend washed oil-bodies in collagenase assay buffer
and add 5 units purified collagenase



Centrifuge to separate out residual oil-bodies



Isolate released peptide from protease treatment
by ammonium sulfate precipitation, column chromatography etc



Perform biological assay on released recombinant peptide

FIG. 4.

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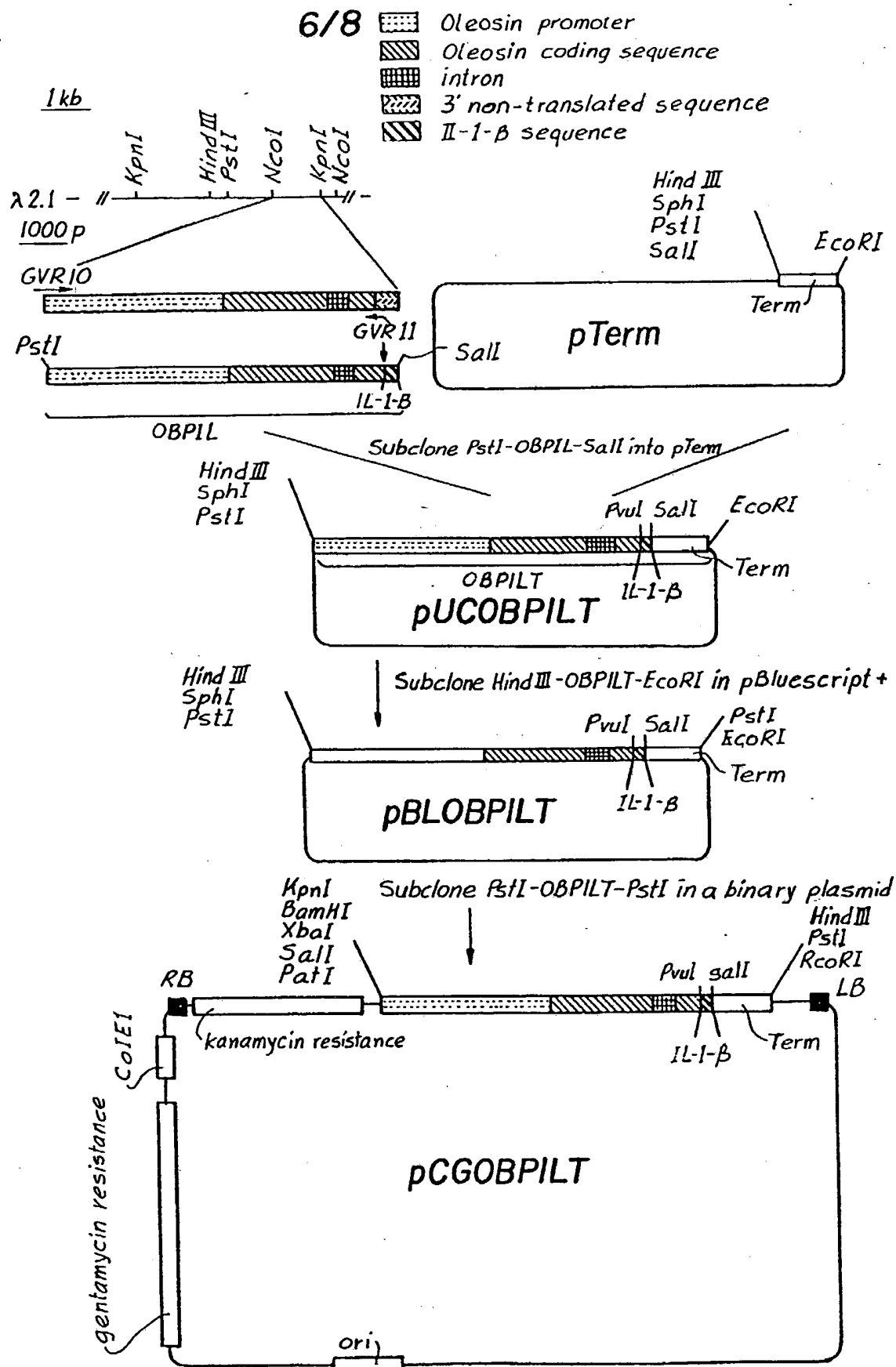


FIG. 5.
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INTERLEUKIN 1- β gvr11 (71-mer)a) A. thaliana oleosin Factor Xa

gly gly gln his thr thr ala ile glu gly arg val gln
 gly glu glu ser asn asp lys OCH val asp

GGT GGC CAG CAC ACT ACT GCT ATC GAA GGG AGA GTT CAG

PvuI

GGA GAA GAA TCT AAC GAC AAG TAA GTC GAC GG

Sali

b) 3'CCA CCG GTC GTG TGA TGA CGC TAG CTT CCC TCT CAA GTC
 [.....
 CCT CTT CTT AGA TTG CTG TTC ATT CAG CTG CC 5'
]

FIG.6.

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PstI

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CACTGCAGGAACCTCTCTGGTAAGCTAGCTCCACTCCCCAGAAACAACCGGCGCCAAATTGCC
GGAATTGCT
GACCTGAAGACGGAACATCATCGTCGGGTCTTGGGCGATTGCGGCGGAAGATGGGTCAGCT
TGGGCTTGAG
GACGAGACCCGAATCGAGTCTGTTGAAAGGTTGTTTCATTGGGATTGTATACGGAGATTGGTC
GTCGAGAGG
TTTGAGGGAAAGGACAAATGGGTTTGGCTCTGGAGAAAGAGAGTGCGGCTTTAGAGAGAGAA
TTGAGAGGTT
TAGAGAGAGATGCGGCGCGGATGACGGGAGGAGAGACGACGAGGACCTGCATTATCAAAGCA
GTGACGTGGT
GAAATTTGGAACTTTAAAGAGGCAGATAGATTTATTATTTGTATCCATTTTCTTCATTGTTT
TAGAATGTCG
CGGAACAAATTTTAAACTAAATCCTAAATTTTCTAATTTTGTGTTGCCAATAGTGGATATGT
GGGCCGTATA
GAAGGAATCTATTGAAGGCCCAAACCCATACTGACGAGCCCAAAGGTTTCGTTTTGCGTTTTA
TGTTTTCGGTT
CGATGCCAACGCCACATTCTGAGCTAGGCCAAAAACAAACGTGTCTTTGAATAGACTCCTCT
CGTTAACACA
TGCAGCGGCTGCATGGTGACGCCATTAAACACGTGGCCTACAATTGCATGATGTCTCCATTGA
CACGTGACTT
CTCGTCTCCTTTCTTAATATATCTAACAAACACTCCTACCTCTTCCAAAATATATACACATC
TTTTTGATCA
ATCTCTCATTTCAAATCTCATTCTCTCTAGTAAACAAGAACAAAAAATGGCGGATACAGCT
AGAGGAACCC
ATCACGATATCATCGGCAGAGACCAGTACCCGATGATGGGCCGAGACCGAGACCAGTACCAG
ATGTCCGGAC
GAGGATCTGACTACTCCAAGTCTAGGCAGATTGCTAAAGCTGCAACTGCTGTACAGCTGGT
GGTTCCCTCC
TTGTTCTCTCCAGCCTTACCCTTGTTGGAAGTGTCTATAGCTTTGACTGTTGCAACACCTCTG
CTCGTTATCT
TCAGCCCCAATCCTTGTCCTCCGGCTCTCATCACAGTTGCACTCCTCATCACCGGTTTTCTTTCC
TCTGGAGGGT
TTGGCATTGCCGCTATAACCGTTTTCTCTTGGATTTACAAGTAAGCACACATTTATCATCTT
ACTTCATAAT
TTTGTGCAATATGTGCATGCATGTGTTGAGCCAGTAGCTTTGGATCAATTTTTTTGGTCGAA
TAACAAATGT
AACAAATAAGAAATTGCAAATCTAGGGAACATTTGGTTAACTAAATACGAAATTTGACCTAG
CTAGCTTGAA
TGTGTCTGTGTATATCATCTATATAGGTAAAAATGCTTGGTATGATACCTATTGATTGTGAAT
AGGTACGCAA
CGGGAGAGCACCCACAGGGATCAGACAAGTTGGACAGTGCAAGGATGAAGTTGGGAAGCAAA
GCTCAGGATC
TGAAAGACAGAGCTCAGTACTACGGACAGCAACATACTGGTTGGGAACATGACCGTGACCGT
ACTCGTGGTG
GCCAGCACACTACTGCGATCGAAGGGAGAGTTTCAGGGAGAAGAATCTAACGACAAGTAAGTC
GACTCTAG
ACGGATCTCCCgatcggttcaaacatttggcaataaagtttcttaagattgaatcctgttgcc
ggtccttgca
tgattatcatataatttctgttgaattacgttaagcatgtaataattaacatgtaatgcatg
acgttatatta
tgagatgggtttttatgattagagtcccgcaattatacatttaatacgcgatagaaaacaaa
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caaactaggataaattatcgcgcgcggtgtcatctatgttactagatcGGAATTC

EcoRI

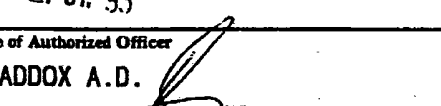
FIG. 7.

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INTERNATIONAL SEARCH REPORT

International Application No

PCT/CA 92/00161

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ⁶		
According to International Patent Classification (IPC) or to both National Classification and IPC		
Int.Cl. 5 C12N15/29; C12N5/10;	C12N15/62; A01H5/00	C12N15/82; C12P21/02
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁷		
Classification System	Classification Symbols	
Int.Cl. 5	C12N ; C12P ; A01H	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ⁸		
III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹		
Category ¹⁰	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
X Y X Y	EMBL Nucleic Acid Sequence Database Accession No. X62353 20-2-1992 Release 31 A. thaliana gene for oleosin --- PLANT MOLECULAR BIOLOGY. vol. 18, no. 6, April 1992, DORDRECHT, THE NETHERLANDS. pages 1177 - 1179 VAN ROOIJEN, G.J.H. 'Nucleotide sequence of an Arabidopsis thaliana oleosin gene' see the whole document --- <div style="text-align: right;">-/--</div>	1,2,4-16 32,33,35 1,2,4-16 32,33,35
<div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <p>¹⁰ Special categories of cited documents :</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="width: 45%;"> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&" document member of the same patent family</p> </div> </div>		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search <div style="text-align: center;">14 DECEMBER 1992</div>	Date of Mailing of this International Search Report <div style="text-align: center;">22.01.93</div>	
International Searching Authority <div style="text-align: center;">EUROPEAN PATENT OFFICE</div>	Signature of Authorized Officer <div style="text-align: center;">MADDOX A.D. </div>	

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category ^a	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
Y	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA vol. 88, July 1991, WASHINGTON US pages 6181 - 6185 LEE, W.S., ET AL. 'Maize oleosin is correctly targeted to seed oil bodies in Brassica napus transformed with the maize oleosin gene' see the whole document	32,33,35
A	J. EXP. BOT., SUPPL., MEETING HELD APRIL 7-12, 1991. vol. 42, no. 238, 1991, page 47 BATCHELDER, C., ET AL. 'Molecular genetics of oil body membrane proteins in Brassica napus and Arabidopsis' see abstract P8.58	1-37
A	BIOTECHNOLOGY vol. 7, no. 9, September 1989, NEW YORK US pages 929 - 932 VANDEKERCKHOVE, J., ET AL. 'Enkephalins produced in transgenic plants using modified 2S seed storage proteins' see the whole document	16,17, 21,23, 28,37
A	PLANT PHYSIOLOGY. vol. 96, 1991, ROCKVILLE, MD, USA. pages 1395 - 1397 LEE, K., ET AL. 'Genomic nucleotide sequence of a Brassica napus 20-kilodalton oleosin gene' see the whole document	1,2,4-16
A	BIOLOGICAL ABSTRACTS vol. 91 , 1991, Philadelphia, PA, US; abstract no. 84634, MURPHY, D.J., ET AL. 'A class of amphipathic proteins associated with lipid storage bodies in plants possible similarities with animal serum apolipoproteins' see abstract & BIOCHIM. BIOPHYS. ACTA vol. 1088, no. 1, 1991, pages 86 - 94	4-9